

4-(Bromomethyl)-2-(methylthio)pyrimidine (26): yield 30%; an unstable oil; $^1\text{H NMR}$ δ 2.62 (s, Me), 4.40 (s, CH_2Br), 7.17 (d, $J = 5$ Hz, 5-H), 8.58 (d, $J = 5$ Hz, 6-H); HRMS m/z (M^+) calcd 217.9514, obsd 217.9521.

5-Bromo-4-(bromomethyl)-2-(methylthio)pyrimidine (27): yield 7%; mp 62-64 °C; $^1\text{H NMR}$ δ 2.60 (s, Me), 4.53 (s, CH_2Br), 8.63 (s, 6-H). Anal. Calcd for $\text{C}_8\text{H}_8\text{Br}_2\text{N}_2\text{S}$: C, 24.18; H, 2.03; N, 9.40. Found: C, 24.27; H, 2.02; N, 9.48.

Oxidation of 24 with Br_2 in AcOH-AcONa. A solution of 24 (0.33 g, 2.36 mmol) and sodium acetate (3.8 g) in acetic acid (8 mL) was heated at 70 °C and treated dropwise within 1 h with a solution of bromine (1.23 g, 7.67 mmol) in acetic acid (7 mL). After an additional 10 min the mixture was cooled, diluted with water (10 mL), neutralized with solid NaHCO_3 , and then extracted with CHCl_3 (6×10 mL).²¹ Chromatography on silica gel (CH_2Cl_2 -AcOEt (8:2)) gave sulfone²² 29 (0.065 g, 16%), which was eluted first, and sulfoxide 28 (0.20 g, 54%). Treatment of 28 with bromine under the same conditions gave 29.

4-Methyl-2-(methylsulfinyl)pyrimidine (28): an oil; $^1\text{H NMR}$ δ 2.70 (s, 4-Me), 3.00 (s, S(O)Me), 7.42 (d, $J = 5$ Hz, 5-H), 8.82 (d, $J = 5$ Hz, 6-H). Anal. Calcd for $\text{C}_8\text{H}_8\text{N}_2\text{OS}$: C, 46.13; H, 5.16; N, 17.94. Found: C, 45.77; H, 5.19; N, 17.65.

4-Methyl-2-(methylsulfonyl)pyrimidine (29): mp 38-40 °C; $^1\text{H NMR}$ δ 2.73 (s, 4-Me), 3.42 (s, SO_2Me), 7.58 (d, $J = 5$ Hz, 5-H), 8.88 (d, $J = 5$ Hz, 6-H). Anal. Calcd for $\text{C}_8\text{H}_8\text{N}_2\text{O}_2\text{S}$: C, 41.84; H, 4.68; N, 16.27. Found: C, 41.54; H, 4.60; N, 15.98.

Reactions of (Bromomethyl)pyrimidines 12, 13, and 16 with DMSO. A solution of 12, 13, or 16 (1 mmol) in anhydrous DMSO (2 mL) was allowed to stand at 23 °C for 24 h and then treated with water (1 mL). The mixture was extracted with ether (5×10 mL), and the extract was dried (Na_2SO_4) and concentrated.

(28) Sulfone 29 is mentioned in: Gefenas, V. *Proceedings of the Conference of Young Chemists*; Vilnius State University: Vilnius, USSR, 1980; p 87; *Chem. Abstr.* 1981, 94, 156861h).

The oxidation of 12 in the presence of NaHCO_3 ¹⁸ (0.5 g) was conducted in a similar manner. Crude products were purified by chromatography.

4-Methyl-2-(methylthio)-5-pyrimidinecarbaldehyde (15): yield 63%; mp 67-69 °C (lit.¹⁷ mp 63-64 °C); $^1\text{H NMR}$ spectrum was identical with that reported.¹⁷

5-Methyl-2-(methylthio)-4-pyrimidinecarbaldehyde (17): yield 78%; mp 56-57 °C; $^1\text{H NMR}$ δ 2.45 (s, 5-Me), 2.60 (s, SMe), 8.75 (s, 6-H), 9.93 (s, CHO); IR (neat) ν 1719 (C=O) cm^{-1} . Anal. Calcd for $\text{C}_7\text{H}_8\text{N}_2\text{OS}$: C, 49.98; H, 4.79; N, 16.66. Found: C, 50.07; H, 4.79; N, 16.58.

5-(Hydroxymethyl)-4-methyl-2-(methylthio)pyrimidine (18a): yield 36%; an oil; $^1\text{H NMR}$ δ 2.48 (s, 4-Me), 2.57 (s, SMe), 3.70 (br s, OH), 4.67 (s, CH_2), 8.38 (s, 6-H). Compound 18a was transformed into a crystalline *p*-nitrobenzoyl derivative 18b as described.¹⁴ mp 149-150 °C (lit.¹⁴ mp 148.5-149.5 °C).

5-(Hydroxymethyl)-2-(methylthio)-4-pyrimidinecarbaldehyde (19a) and 6-(Methylthio)-1,3-dihydrofuro[3,4-*d*]pyrimidin-1-ol (19b): yield 74%; mp 136-137 °C (lit.¹⁹ mp 135-136 °C); IR (KBr) ν 3283 (OH) cm^{-1} ; no C=O absorption; IR (CHCl_3) ν 1735 (C=O) cm^{-1} . The two forms exist in CDCl_3 solution in the ratio 19a:19b = 1:3. $^1\text{H NMR}$ (400 Hz) for 19a: δ 2.65 (s, SMe), 3.00 (t, $J = 6$ Hz, OH), 4.84 (d, $J = 6$ Hz, 5- CH_2), 8.80 (s, 6-H), 10.04 (s, CHO). $^1\text{H NMR}$ (400 Hz) for 19b: δ 2.61 (s, SMe), 3.51 (d, $J = 6$ Hz, OH), 5.05 and 5.26 (2d, $J = 13$ Hz, 3a- CH_2), 6.33 (s, 7a-CH), 8.56 (s, 4-H).

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Regioselective Functionalization of *N*-Phthaloyl-Substituted Amino Acid and Peptide Derivatives

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The free-radical reactions of a range of amino acid derivatives with *N*-bromosuccinimide are described. The products and relative rates of these reactions indicate that the α -position of an *N*-phthaloyl-substituted α -amino acid derivative is much less reactive than that of a corresponding *N*-acyl amino acid derivative toward hydrogen atom transfer. This is attributed to the proactive effects of acylamino and carboxyl substituents, in contrast to the counteractive effects of phthalimido and carboxyl groups. The reactions exemplify procedures for the regiocontrolled functionalization of amino acid and peptide derivatives.

Introduction

Hydrogen atom transfer reactions of *N*-acyl α -amino acid derivatives generally favor formation of α -carbon-centered radicals.¹ These radicals are resonance stabilized by the combined action of an electron-releasing amido substituent and an electron-withdrawing carboxy substituent, and they may be classified as captodative,² me-

rostabilized,³ or push-pull-stabilized⁴ radicals. Amido-, carboxy-substituted radicals have been identified in proteins upon irradiation⁵ and are thought to be intermediates in the photoalkylation⁶ and carboxylation⁷ of peptides.

(3) Baldock, R. W.; Hudson, P.; Katritzky, A. R.; Soti, F. *Heterocycles* 1973, 1, 67. Baldock, R. W.; Hudson, P.; Katritzky, A. R.; Soti, F. *J. Chem. Soc., Perkin Trans. 1* 1974, 1422.

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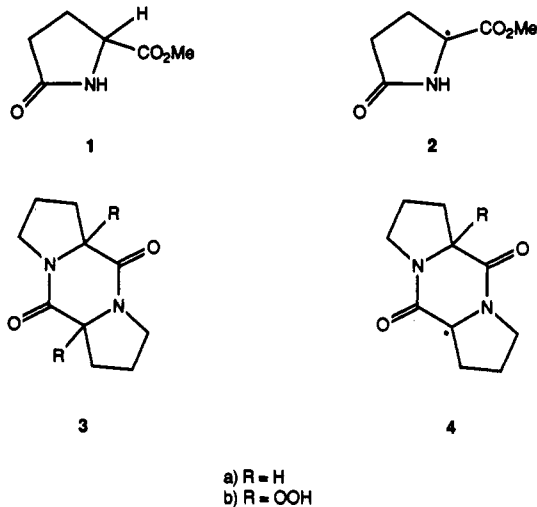
(5) Henrikson, T.; Melo, T. B.; Saxebol, G. *Free Radicals in Biology*; Pryor, W. A., Ed.; Academic Press: New York, 1976; Vol. 2, p 213.

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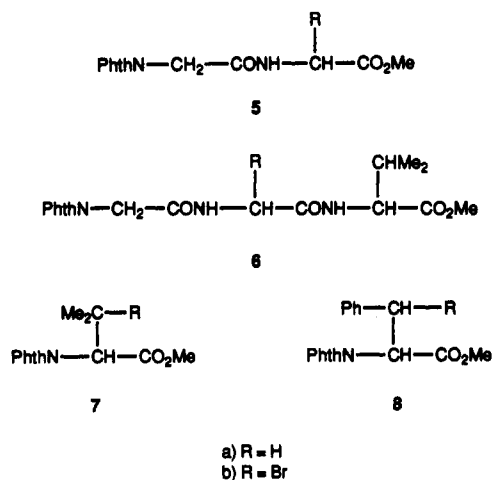
(1) For a review of the formation of α -carbon-centered radicals in reactions of amino acid derivatives, see: Easton, C. J. *Advances in Detailed Reaction Mechanisms*; Coxon, J. M., Ed.; JAI Press: Greenwich, CT, 1991; Vol. 1, p 83.

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Other examples of their generation include the formation and dimerization of the radical 2 on irradiation of a mixture of methyl pyroglutamate (1) and di-*tert*-butyl peroxide,⁸ and the oxidation of 3a to give the diperoxide 3b, presumably via 4a and 4b.⁹

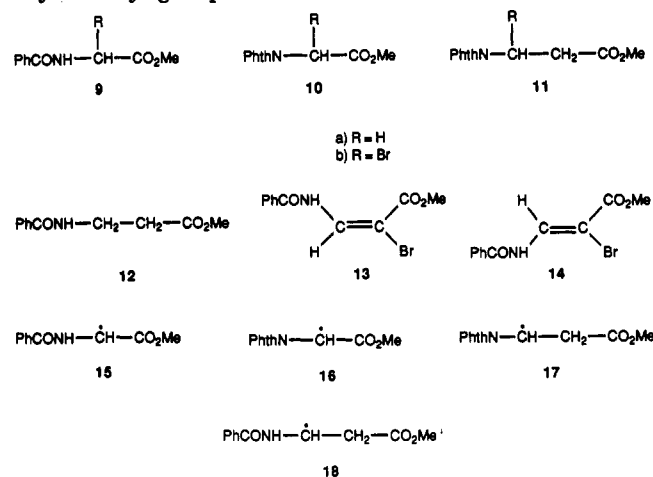


Consistent with this general trend, reactions of *N*-acyl α -amino acid derivatives with *N*-bromosuccinimide (NBS) proceed via formation of the corresponding α -carbon-centered radicals.^{10,11} In direct contrast, however, formation of α -carbon-centered radicals from *N*-phthaloyl-substituted amino acid derivatives is strongly disfavored.¹² The extent of this effect is reflected in the regiospecific reactions of 5a–8a to give the corresponding bromides 5b–8b. In this report we describe the reactions of NBS with a range of amino acid derivatives, chosen to examine the cause of the contrasting effects of the *N*-phthaloyl and *N*-acyl substituents and to further illustrate the exploitation of those effects in the regioselective functionalization of amino acid and peptide derivatives.



Results and Discussion

To investigate the contrasting effects of *N*-phthaloyl and *N*-acyl substituents on the formation of α -carbon-centered radicals in reactions of α -amino acid derivatives, reactions of the methyl esters of *N*-benzoylglycine (9a), *N*-phthaloylglycine (10a), *N*-phthaloyl- β -alanine (11a), and *N*-benzoyl- β -alanine (12) with NBS were investigated and compared. The β -alanine derivatives 11a and 12, which have the phthalimido and benzamido substituents spacially separated from the methoxycarbonyl group, were chosen to examine the relative effects of these substituents acting individually, while the glycine derivatives 9a and 10a were selected to study the effects of the benzamido and phthalimido substituents in combination with the methoxycarbonyl group.



The reactions of 9a and 11a with NBS (1 equiv) in carbon tetrachloride gave the corresponding bromides 9b and 11b. Similar treatment of 12 gave a mixture of the α,β -dehydro- β -alanine derivatives 13 and 14 and unreacted starting material, while with excess NBS (2 equiv) all of the starting material was consumed. The reaction of 9a was complete in 0.25 h, whereas the reactions of 11a and 12 required 2 h for complete reaction. The reaction of 10a to give 10b was much less efficient, requiring excess NBS (2 equiv) and a reaction time of 48 h for 50% conversion. This qualitative comparison of the reactivity of 9a, 10a, 11a, and 12 with NBS is reflected in the relative rates of reaction of the amino acid derivatives, which were determined in competitive experiments using, for example, equimolar mixtures of 9a and 12, 10a and 11a, and 11a and 12. Aliquots of crude reaction mixtures were easily and conveniently analyzed, by ¹H NMR spectroscopy, for consumption of starting materials and formation of products. In the competitive reaction of 9a and 12 there was no evidence for any reaction of 12 until reaction of 9a to give 9b appeared to be complete. Similarly, with mixtures of 10a and 11a, and 11a and 12, 11a reacted to the exclusion of 10a, and 12 reacted to the exclusion of 11a, within the limits of detection by ¹H NMR spectroscopy. From these experiments a conservative estimate for the relative rates of reaction of 9a and 12, 12 and 11a, and 11a and 10a is 10:1 in each case.

The products of treatment of 9a, 10a, 11a, and 12 with NBS indicate that the reactions involve formation of the corresponding radicals 15–18, with subsequent bromine incorporation. In the reaction of 12, subsequent elimination of hydrogen bromide, bromine addition, and hydrogen bromide elimination afford 13 and 14. The regioselectivity of hydrogen atom transfer from 12 can be deduced from the greater relative rate of reaction of 12 compared to 11a and the regioselectivity of reaction of 11a. Reaction at the

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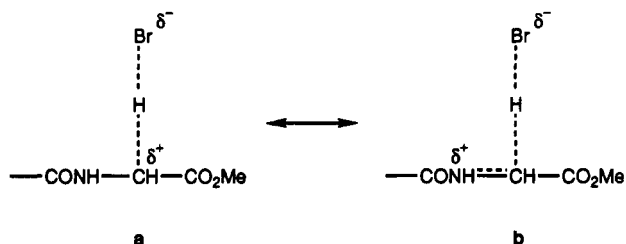


Figure 1. (a) Development of charge in the transition state for hydrogen atom abstraction by bromine atom and (b) delocalization of that charge by an amido substituent.

methylene adjacent to the methoxycarbonyl substituent in 12 would be inconsistent with the greater relative rate of reaction of 12 compared to 11a and the reaction of 11a at the methylene substituted with the phthalimido group in preference to reaction adjacent to the methoxycarbonyl substituent.

On this basis, the relative rates of reaction of 9a, 10a, 11a, and 12 reflect the comparative ease of formation of the corresponding radicals 15–18. The faster rate of reaction of 12 compared to 11a can be attributed to the greater resonance stabilization provided by the benzamido substituent in 18 than by the phthalimido substituent in 17. From the greater relative rate of formation of 15 compared to 18, the effect of the methoxycarbonyl group in combination with the benzamido substituent is proactive, whereas the slower relative rate of formation of 16 compared to 17 shows that the effect of the methoxycarbonyl group in combination with the phthalimido substituent is counteractive. A radical formed adjacent to a methoxycarbonyl substituent is resonance stabilized, but formation of such a radical by hydrogen transfer to a bromine atom is often disfavored by a polar effect, involving the inductive interaction between the electron-deficient center of the substituent and that developing in the transition state at the site of hydrogen abstraction (Figure 1a).¹³ The proactive nature of the methoxycarbonyl group in combination with the benzamido substituent may be attributed to the ability of the benzamido substituent to delocalize charge developed in the transition state (Figure 1b). The resulting diminution of the polar effect of the methoxycarbonyl substituent leads to the enhanced rate of formation of the resonance-stabilized radical 15. The charge delocalization provided by the phthalimido substituent is less than that of the benzamido group, to the extent that this effect in the reaction of 10a is outweighed by steric effects, resulting in the counteractive effect of the methoxycarbonyl group in combination with the phthalimido substituent. The steric effects will arise from interactions of the methoxycarbonyl and phthalimido substituents in 10a with the hydrogen-abstracting species, and from interactions between the methoxycarbonyl and phthalimido substituents preventing the radical 16 from adopting planar conformations in which there is maximum delocalization of the unpaired spin density (Figure 2). These interactions will be greater than those between the methoxycarbonyl and benzamido substituents in 9a and 15.

In our initial report,¹² the contrasting effects of *N*-phthaloyl and *N*-acyl substituents were illustrated with

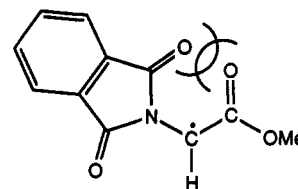
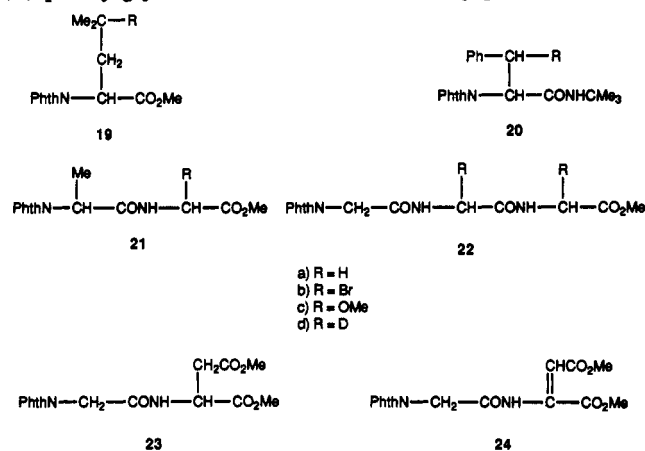


Figure 2. Nonbonding interactions associated with planar conformations of the radical 16.

the reactions of 5a–8a to give 5b–8b, respectively. The reactions of 19a–22a and 23 with NBS further demonstrate the extent of the counteractive effect of a carboxyl substituent in combination with the phthalimido substituent, to disfavor reaction at the α -position of amino acid derivatives. Treatment of the derivatives of leucine 19a and phenylalaninamide 20a with NBS (1 equiv) gave the corresponding bromides 19b and 20b, each in high yield. The reaction was repeated with the *S* isomer of 20a, and the bromide 20b thus obtained was treated with tributyltin hydride. The reduced product 20a retained the homochirality of the starting material, as shown by comparison with a racemic sample using an HPLC column with (*S*)-phenylglycine as the chiral stationary phase.



The reaction of the peptide derivative 21a with NBS (1 equiv) in dichloromethane gave the bromide 21b, which was characterized by conversion to the methoxyglycine derivative 21c and the deuterated peptide derivative 21d. Mass spectrometric analysis indicated that the deuterium incorporation of 90% in 21d was regiospecific. The methoxyglycine derivative 21c was obtained as a 1:1 mixture of diastereomers, as determined by ¹H NMR spectroscopy. The tripeptide derivative *N*-phthaloylglycylglycylglycine methyl ester (22a) was only sparingly soluble in dichloromethane, and its reaction with NBS had to be conducted in dilute solution. Under those conditions some of the NBS was consumed through reaction with the solvent. Thus, an excess of NBS (4 equiv) was required to produce the dibromide 22b. Treatment of the crude dibromide 22b with tributyltin deuteride gave 22d, in 55% yield based on 22a, with 90% and 95% deuterium incorporation at the C-terminal and nonterminal glycine residues, respectively. The reaction of *N*-phthaloylglycyl-aspartic acid dimethyl diester (23) with NBS gave an 82% yield of the α,β -dehydroaspartate derivative 24, which was assumed to have *Z* stereochemistry on the basis of the tendency of dehydro amino acid derivatives to favor this configuration.¹⁴

In summary, the reactions of 5a–11a, 12, 19a–22a, and 23 illustrate the contrasting effects of *N*-phthaloyl and

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N-acyl substituents on the regioselectivity of reactions of amino acid and peptide derivatives with NBS. They exemplify procedures for the functionalization of amino acid derivatives and for the regiocontrolled halogenation of peptide derivatives.

Experimental Section

General. Melting points are uncorrected. ^1H NMR spectra were recorded as dilute solutions in deuteriochloroform on a Bruker CXP-300 spectrometer. Chemical shifts (δ) are reported in parts per million downfield from tetramethylsilane. Electron-impact mass spectra were recorded on an AEI MS-3010 spectrometer. Microanalyses were performed by the Canadian Microanalytical Service Ltd., New Westminster, British Columbia, Canada. HPLC was performed by using a Waters Model 501 solvent delivery system and a U6K injector with a Waters Model 481 absorbance detector. Preparative chromatography was carried out on a Chromatotron 7924T instrument (Harrison Research, Palo Alto/TC Research, Norwich, CA) by using Merck silica gel 60 PF₂₅₄, eluting with a gradient of light petroleum/dichloromethane/ethyl acetate. Light petroleum refers to the fraction with bp 55–65 °C. Carbon tetrachloride and dichloromethane were purified by fractional distillation and stored over 4A molecular sieves. NBS was recrystallized from water and dried under reduced pressure before use. A Philips MLU 300-W (220–240-V) ultraviolet lamp was used as the light source in reactions of NBS. Tributyltin deuteride was prepared by reduction of tributyltin chloride with lithium aluminum deuteride.¹⁵ N-Benzoylglycine methyl ester (9a),¹⁶ N-phthaloylglycine methyl ester (10a),¹⁷ N-phthaloyl- β -alanine methyl ester (11a),¹⁸ N-benzoyl- β -alanine methyl ester (12),¹⁹ N-phthaloylleucine methyl ester (19a),²⁰ (S)- and (RS)-N-(tert-butyl)-N $^{\alpha}$ -phthaloylphenylalaninamide (20a), N-phthaloylalaninylglycine methyl ester (21a),²¹ N-phthaloyl-glycylglycylglycine methyl ester (22a),²¹ and N-phthaloylglycyl-aspartic acid dimethyl diester (23) were prepared by using standard procedures.

N-Benzoyl- α -bromoglycine Methyl Ester (9b). A mixture of the glycine derivative 9a (200 mg, 1 mmol) and NBS (180 mg, 1 mmol) in carbon tetrachloride (20 mL) was heated at reflux under nitrogen for 0.25 h, with reaction initiated by irradiation with a 300-W ultraviolet lamp; then the reaction mixture was cooled, filtered, and concentrated under reduced pressure to give crude 9b;²² ^1H NMR 3.93 (3 H, s), 6.65 (1 H, d, J = 8 Hz), 7.30–7.90 (6 H, m).

N-Phthaloyl- α -bromoglycine Methyl Ester (10b). Treatment of 10a with NBS as described above for the preparation of 9b, except that excess NBS (2 equiv) was used and the reaction mixture was heated at reflux for 48 h, gave a mixture of 10a and 10b in a ratio of ca. 1:1. Chromatography of the mixture on silica gave 10b in 27% yield: mp 116–118 °C; ^1H NMR 3.88 (3 H, s), 6.68 (1 H, s), 7.80–8.00 (4 H, m); MS m/e 240, 238, 218, 160; MS m/e 237.9508 (M^+ – COOMe). Calcd for $\text{C}_9\text{H}_9\text{BrNO}_3$: 237.9504.

N-Phthaloyl- β -bromo- β -alanine Methyl Ester (11b). Treatment of 11a with NBS as described above for the preparation of 9b, except that the reaction mixture was heated at reflux for 2 h, gave 11b in 84% yield: mp 95–97 °C; ^1H NMR 3.64 (1 H, dd, J = 7 and 17 Hz), 3.70 (3 H, s), 3.89 (1 H, dd, J = 8 and 17 Hz), 6.59 (1 H, dd, J = 7 and 8 Hz), 7.20–7.90 (4 H, m); MS m/e 313, 311, 282, 280, 254, 252, 240, 238, 232, 200, 173, 172, 160. Anal. Calcd for $\text{C}_{12}\text{H}_{10}\text{BrNO}_4$: C, 46.2; H, 3.2; N, 4.5. Found: C, 46.4; H, 3.2; N, 4.6.

Reaction of N-Benzoyl- β -alanine Methyl Ester (12) with NBS. Treatment of 12 with NBS as described above for the

preparation of 9b, except that the reaction mixture was heated at reflux for 2 h, gave an oil that contained unreacted starting material and the dehydro- β -alanine derivatives 13 and 14, in a ratio of ca. 3:2:1, as determined by ^1H NMR spectroscopic analysis. The components were separated by chromatography of the mixture on silica, to give 13 and 14 in yields of 32% and 15%, respectively. The stereochemistry of 13 and 14 was assigned on the basis of an X-ray crystal structure determination.²³

Methyl (E)-3-benzamido-2-bromoacrylate (13): mp 98–99 °C; ^1H NMR 3.89 (3 H, s), 7.40–7.90 (5 H, m), 8.38 (1 H, br d, J = 12 Hz), 8.60 (1 H, d, J = 12 Hz). Anal. Calcd for $\text{C}_{11}\text{H}_{10}\text{BrNO}_3$: C, 46.5; H, 3.5. Found: C, 46.5; H, 3.5.

Methyl (Z)-3-benzamido-2-bromoacrylate (14): mp 122.5–123.5 °C; ^1H NMR 3.80 (3 H, s), 7.40–7.90 (5 H, m), 8.20 (1 H, d, J = 12 Hz), 11.30 (1 H, br d, J = 12 Hz). Anal. Calcd for $\text{C}_{11}\text{H}_{10}\text{BrNO}_3$: C, 46.5; H, 3.5. Found: C, 46.5; H, 3.7.

N-Phthaloyl- γ -bromoleucine Methyl Ester (19b). Treatment of 19a with NBS as described above for the preparation of 9b gave 19b in 82% yield: mp 63–64 °C; ^1H NMR 1.76 (3 H, s), 1.84 (3 H, s), 2.85 (2 H, m), 3.74 (3 H, s), 5.25 (1 H, dd, J = 4.5 and 8 Hz), 7.75–7.90 (4 H, m); MS m/e 296, 294, 274, 214. Anal. Calcd for $\text{C}_{16}\text{H}_{16}\text{BrNO}_4$: C, 50.8; H, 4.6; N, 4.0. Found: C, 50.9; H, 4.6; N, 4.0.

N-tert-Butyl-N $^{\alpha}$ -phthaloyl- β -bromophenylalaninamide (20b). Treatment of (RS)-20a with NBS as described above for the preparation of 9b gave 20b as a 1:1 mixture of diastereomers, which were separated by fractional crystallization from 2-propanol/light petroleum. One diastereomer was obtained as needle-shaped crystals in 41% yield: mp 199–200 °C; ^1H NMR 1.03 (9 H, s), 5.28 (1 H, d, J = 12 Hz), 5.85 (1 H, br s), 6.25 (1 H, d, J = 12 Hz), 7.30–8.00 (9 H, m); MS m/e 430, 428. Anal. Calcd for $\text{C}_{21}\text{H}_{21}\text{BrN}_2\text{O}_3$: C, 58.8; H, 4.9; N, 6.5. Found: C, 58.8; H, 4.9; N, 6.6. The other diastereomer was isolated as pale-yellow granular crystals in 39% yield: mp 213–213.5 °C; ^1H NMR 1.43 (9 H, s), 5.32 (1 H, d, J = 11.5 Hz), 6.04 (1 H, d, J = 11.5 Hz), 6.50 (1 H, br s), 7.20–7.80 (9 H, m); MS m/e 430, 428. Anal. Calcd for $\text{C}_{21}\text{H}_{21}\text{BrN}_2\text{O}_3$: C, 58.8; H, 4.9; N, 6.5. Found: C, 58.5; H, 5.0; N, 6.8.

Reaction of (S)-N-tert-Butyl-N $^{\alpha}$ -phthaloylphenylalaninamide (20a) with NBS and Tributyltin Hydride. Treatment of (S)-20a with NBS as described above for the reaction of (RS)-20a gave a 1:1 mixture of the diastereomers of the bromide (20b). A sample of this mixture of the bromides (20b) (100 mg, 0.23 mmol) was treated with tributyltin hydride (67 mg, 0.23 mmol) in benzene (5 mL), at reflux under nitrogen for 4 h. The product (20a) (71 mg, 88%) showed only one component, with a retention time of 21 min, on HPLC analysis using a Regis Pirkle covalent (S)-phenylglycine column (25 cm \times 4.6 mm), eluting with a gradient of light petroleum/2-propanol (1 mL/min, 5–40% over 40 min). By comparison, under the same conditions (RS)-20a resolved into two components, with retention times of 21 and 22 min.

N-Phthaloylalaninyl- α -methoxyglycine Methyl Ester (21c). The peptide derivative 21a (0.7 mmol) was treated with NBS (125 mg, 0.7 mmol) in dichloromethane (25 mL), at reflux under nitrogen for 0.5 h, with reaction initiated by irradiation with a 300-W ultraviolet lamp. The reaction mixture was cooled and filtered, and the filtrate was concentrated under reduced pressure to give the crude bromide 21b (^1H NMR 6.56, d, J = 10 Hz). Alternatively, methanol (0.5 mL) was added to the filtrate and the mixture was stirred at room temperature for 2 h; then it was concentrated under reduced pressure. The residual oil was chromatographed on silica and crystallized from dichloromethane/light petroleum to give the methoxyglycine derivative 21c in 70% yield: mp 134–139 °C; ^1H NMR 1.72 (1.5 H, d, J = 7.5 Hz), 1.74 (1.5 H, d, J = 7.5 Hz), 3.45 (1.5 H, s), 3.47 (1.5 H, s), 3.78 (1.5 H, s), 3.79 (1.5 H, s), 5.00 (1 H, m), 5.56 (0.5 H, d, J = 9 Hz), 5.58 (0.5 H, d, J = 9 Hz), 6.93 (0.5 H, br d, J = 9 Hz), 6.98 (0.5 H, br d, J = 9 Hz), 7.70–7.90 (4 H, m); MS m/e 261, 202, 175, 174. Anal. Calcd for $\text{C}_{15}\text{H}_{16}\text{N}_2\text{O}_6$: C, 56.3; H, 5.0; N, 8.7. Found: C, 56.4; H, 5.0; N, 8.7.

N-Phthaloylalaninyl- α -deuteriogylicine Methyl Ester (21d). Tributyltin deuteride (310 mg, 1.07 mmol) was added to a crude,

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filtered reaction mixture of the bromide 21b, prepared from 21a (0.7 mmol) as described above. The mixture was stirred at room temperature for 2 h; then it was concentrated under reduced pressure. The residual oil was chromatographed on silica and crystallized from dichloromethane/light petroleum to give 21d in 70% yield: mp 145–146 °C; $^1\text{H NMR}$ 1.73 (3 H, d, $J = 7.5$ Hz), 3.73 (3 H, s), 4.05 (1.1 H, d, $J = 5$ Hz), 4.98 (1 H, q, $J = 7.5$ Hz), 6.71 (1 H, br d, $J = 5$ Hz), 7.70–7.85 (4 H, m); MS m/e 291 (90% $^2\text{H}_1$), 232 (90% $^2\text{H}_1$), 202 (0% $^2\text{H}_1$).

N-Phthaloylglycyl- α -deuteriogylicyl- α -deuteriogylicine Methyl Ester (22d). Addition of tributyltin deuteride (530 mg, 1.8 mmol) to a crude, filtered reaction mixture of 22b, prepared from 22a (200 mg, 0.6 mmol) and NBS (430 mg, 2.4 mmol) in dichloromethane (200 mL), as described above for the preparation of 21b, gave 22d (0.11 g, 55%): mp 230–231 °C; $^1\text{H NMR}$ 3.69 (3 H, s), 3.86 (1.1 H, d, $J = 5$ Hz), 3.90 (1.05 H, d, $J = 6$ Hz), 4.37 (2 H, s), 7.70–7.90 (4 H, m), 8.14 (1 H, br d, $J = 6$ Hz), 8.51 (1 H, br d, $J = 5$ Hz); MS m/e 335 (90% $^2\text{H}_2$, 5% $^2\text{H}_1$), 246 (95% $^2\text{H}_1$), 218 (95% $^2\text{H}_1$), 188 (0% $^2\text{H}_1$).

N-Phthaloylglycyl-(Z)- α,β -dehydroaspartic Acid Dimethyl Diester (24). Treatment of 23 (400 mg, 1.15 mmol) with

NBS (205 mg, 1.15 mmol), as described above for the preparation of 21b, gave 24 (330 mg, 82%): mp 175–176 °C; $^1\text{H NMR}$ 3.73 (3 H, s), 3.79 (3 H, s), 4.50 (2 H, s), 5.57 (1 H, s), 7.70–7.90 (4 H, m), 10.50 (1 H, br s); MS m/e 346, 345, 314, 287, 188, 161, 160. Anal. Calcd for $\text{C}_{16}\text{H}_{14}\text{N}_2\text{O}_7$: C, 55.5; H, 4.1; N, 8.1. Found: C, 55.5; H, 4.0; N, 8.1.

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Registry No. 9a, 1205-08-9; 9b, 101649-82-5; 10a, 23244-58-8; 10b, 135395-15-2; 11a, 39739-01-0; 11b, 135395-26-5; 12, 89928-06-3; 13, 129309-14-4; 14, 129309-13-3; 19a, 132785-19-4; 19b, 132785-25-2; (S)-20a, 135395-13-0; (RS)-20a, 135501-56-3; (2S,3R)-20b, 135395-16-3; (2S,3S)-20b, 135395-17-4; (\pm)-(R*,R*)-20b, 135501-57-4; (\pm)-(R*,S*)-20b, 135501-58-5; 21a, 63267-72-1; 21b (diastereomer 1), 135395-18-5; 21b (diastereomer 2), 135395-19-6; 21c (diastereomer 1), 135395-20-9; 21c (diastereomer 2), 135395-21-0; 21d (diastereomer 1), 135395-22-1; 21d (diastereomer 2), 135395-23-2; 22a, 63199-92-8; 22b, 135395-24-3; 22d, 135395-25-4; 23, 135395-14-1; 24, 87358-90-5.

Variculanol: Structure and Absolute Stereochemistry of a Novel 5/12/5 Tricyclic Sesterterpenoid from *Aspergillus varicolor*

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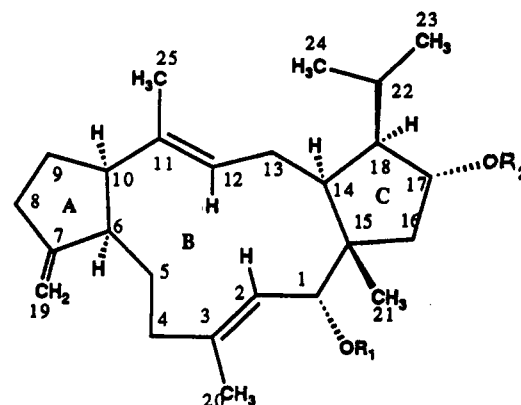
The structure, absolute stereochemistry, and conformation of variculanol, a sesterterpenoid with novel skeleton isolated from *Aspergillus varicolor*, has been described. The molecular structure was deduced from extensive application of 2D NMR methods, in particular HMBC, which established the unambiguous assignment of the novel 5/12/5 ring system. NOEDS measurements were very useful in establishing the relative stereochemistry. The NMR–mandelate method and CD spectral analysis of the 1,17-bis(4-bromobenzoate) were used to determine the absolute stereochemistry. A unique solution-phase conformation was determined from both extensive NOE measurements and MM2 calculations and optimization.

In our search for novel antiparasitic compounds effective against coccidia,¹ we discovered variculanol (1a), a sesterterpenoid having a novel 5/12/5 ring system from *Aspergillus varicolor*. We report herein the isolation, structure, absolute stereochemistry, and solution conformation of variculanol (1a).

A methyl ethyl ketone extract of a solid-state fermentation of *A. varicolor* was partitioned between hexane and methanol–water. Silica gel chromatography of the hexane extract followed by crystallization from CH_3CN afforded granules of variculanol² (1a).

Structure Elucidation. High-resolution EI mass spectral analysis of variculanol (1a) gave the molecular formula $\text{C}_{25}\text{H}_{40}\text{O}_2$ with six double-bond equivalents (DBE), which was corroborated by ^{13}C NMR spectral data (Table I). The IR spectrum of 1a showed hydroxy absorption, which was confirmed by formation of a diacetate (1b, M^+ , 456; IR 1733 cm^{-1}) and a bis(trimethylsilyl ether) (M^+ , 516.3809). Because of the presence of only three double bonds, this molecule must have a tricyclic skeleton.

The 400-MHz ^1H NMR spectrum of 1a in CDCl_3 and $\text{DMSO}-d_6$ (Table I) exhibited some readily assignable



- 1a: $\text{R}_1 = \text{R}_2 = \text{H}$, variculanol
 b: $\text{R}_1 = \text{R}_2 = \text{acetyl}$
 c: $\text{R}_1 = (\text{R})\text{-O-methylmandeloyl}$, $\text{R}_2 = \text{H}$
 d: $\text{R}_1 = \text{H}$, $\text{R}_2 = (\text{R})\text{-O-methylmandeloyl}$
 e: $\text{R}_1 = (\text{S})\text{-O-methylmandeloyl}$, $\text{R}_2 = \text{H}$
 f: $\text{R}_1 = \text{H}$, $\text{R}_2 = (\text{S})\text{-O-methylmandeloyl}$
 g: $\text{R}_1 = 4\text{-bromobenzoyl}$, $\text{R}_2 = \text{H}$
 h: $\text{R}_1 = \text{H}$, $\text{R}_2 = 4\text{-bromobenzoyl}$
 i: $\text{R}_1 = \text{R}_2 = 4\text{-bromobenzoyl}$

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signals such as two vinylic methyls, an angular methyl and a set of methyl doublets, two oxymethines, two exocyclic methylene protons, and a pair of olefinic protons. The